

## Five New $\beta$ -Carboline-Type Alkaloids from *Stellaria dichotoma* var. *lanceolata*

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Five new  $\beta$ -carboline-type alkaloids, dichotomines F–J (**1–5**, resp.), along with nine known compounds, dichotomides I, III, V, and VII (**6–9**, resp.), stellarines A and C (**10–11**, resp.), dichotomine B (**12**), glucodichotomine B (**13**), and 1-acetyl-3-carboxy- $\beta$ -carboline (**14**), were isolated from the roots of Chinese medicinal plant *Stellaria dichotoma* L. var. *lanceolata*. Their structures were determined by chemical and spectroscopic means. Compounds **12** and **13** exhibited moderate cytotoxicity.

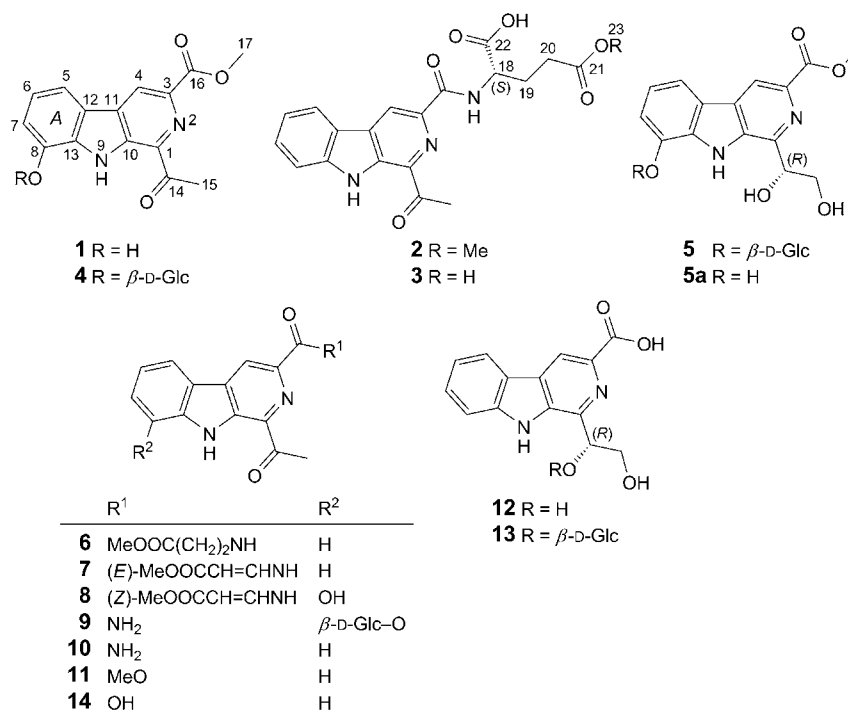
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**Introduction.** – The *Stellaria dichotoma* L. var. *lanceolata* BUNGE (Caryophyllaceae) is distributed in Ningxia and neighboring provinces of China. Its roots are being used as a traditional Chinese medicine (*Yin-Chai-Hu*) to treat fever, consumptive disease, and the infantile malnutrition syndrome [1]. Previous investigations of this species have led to isolation of a series of compounds including flavonoids, sterols, cyclic peptides, neolignans, phenylpropanoids, and  $\beta$ -carboline alkaloids [1–9], some of which displayed various biological properties, such as antifebrile [1–3], antiallergic [3], vasorelaxant [4], cytotoxic [5][6], antibacterial, antifungal, and anti-inflammatory activities [7][10]. As part of our ongoing studies on the chemical constituents of medicinal plants in the Caryophyllaceae [11–13], five new  $\beta$ -carboline alkaloids, dichotomines F–J<sup>1)</sup> (**1–5**, resp.), along with nine known compounds, **6–14**, were isolated from the roots of *S. dichotoma* var. *lanceolata* (Fig. 1). The cytotoxicities of **1–14** against human cancer cell lines Bel7402, SMMC-7721, HCT116, and H460 were evaluated. Here, the isolation, structure elucidation, and cytotoxic activities of these isolates are reported.

**Results and Discussion.** – *Structure Elucidation.* Compound **1** was isolated as yellow powder, presumably being endowed with a N function on the basis of TLC examinations by using *Dragendorff's* reagent. The molecular formula  $C_{15}H_{12}N_2O_4$  was deduced from the *quasi*-molecular-ion peak at  $m/z$  307.0686 ( $[M + Na]^+$ ; calc. 307.0689) in the HR-ESI-MS. The IR spectrum evidenced the presence of OH ( $3373\text{ cm}^{-1}$ ), CO ( $1702\text{ cm}^{-1}$ ), and aromatic groups ( $1437\text{ cm}^{-1}$ ). Absorption maxima in the UV spectrum of **1** were observed at 385, 286, and 236 nm, suggesting the presence of a  $\beta$ -carboline chromophore [14][15].

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<sup>1)</sup> For systematic names, see *Exper. Part*.


 Fig. 1. Structures of **1–14**

The <sup>1</sup>H-NMR ((D<sub>6</sub>)DMSO) spectrum of **1** (Table 1) exhibited signals of a Me group at  $\delta$ (H) 2.83 (s), of a MeO group at  $\delta$ (H) 3.97 (s), a *singlet*, characteristic for a  $\beta$ -carboline skeleton at  $\delta$ (H) 9.11 (s, H-C(4)), and two broad *singlets* at  $\delta$ (H) 10.13 and 11.75 (D<sub>2</sub>O-exchangeable). Moreover, three vicinally coupled aromatic H-atom signals ( $\delta$ (H) 7.93 (*d*, *J* = 8.0), 7.23 (*t*, *J* = 8.0), and 7.05 (*d*, *J* = 7.8)) were indicative of a trisubstituted aromatic ring A within the  $\beta$ -carboline unit. The <sup>13</sup>C-NMR spectrum of **1** (Table 2), along with the information obtained from the HSQC experiment, showed 15 C-atom signals. Besides the signals of a  $\beta$ -carboline alkaloid skeleton, there were those of one Me C-atom at  $\delta$ (C) 25.5, of one MeO C-atom at  $\delta$ (C) 52.2, and of two CO C-atoms at  $\delta$ (C) 201.2 and 165.2, suggesting the presence of ketone and ester CO group. In the HMBC spectrum, a correlation was observed between the signals at  $\delta$ (H) 10.13 (br. *s*) and  $\delta$ (C) 143.4 (C(8)), indicating the presence of a phenolic OH group at C(8) (Fig. 2). Furthermore, the correlations between the signals at  $\delta$ (H) 2.83 (Me(15)), and those at  $\delta$ (C) 135.3 (C(1)) and 201.2 (C(14)), as well as between the signals at  $\delta$ (H) 3.97 (Me(17)), and those at  $\delta$ (C) 135.7 (C(3)) and 165.2 (C(16)) suggested that the Ac and COOMe groups were at C(1) and C(3), respectively (Fig. 2). Thus, the chemical structure of **1** was deduced as shown in Fig. 1 and named dichotomine F.

Compounds **2–5** were also identified as  $\beta$ -carboline derivatives, since they all were found to display the same characteristic UV- and IR-spectroscopic data as compound **1**, and they also showed positive responses toward *Dragendorff's* reagent. Compound **2**

Table 1.  $^1\text{H-NMR}$  Spectroscopic Data for the Isolated  $\beta$ -Carboline Derivatives **1–5** (500 MHz,  $(\text{D}_6)$ DMSO,  $\delta$  in ppm,  $J$  in Hz). Atom numbering as indicated in Fig. 1.

	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>
H–C(4)	9.11 ( <i>s</i> )	9.11 ( <i>s</i> )	9.08 ( <i>s</i> )	9.19 ( <i>s</i> )	8.83 ( <i>s</i> )
H–C(5)	7.93 ( <i>d</i> , $J=8.0$ )	8.46 ( <i>d</i> , $J=8.0$ )	8.44 ( <i>d</i> , $J=7.5$ )	8.17 ( <i>d</i> , $J=8.0$ )	8.06 ( <i>d</i> , $J=7.8$ )
H–C(6)	7.23 ( <i>t</i> , $J=8.0$ )	7.35 ( <i>t</i> , $J=7.8$ )	7.33 ( <i>t</i> , $J=7.5$ )	7.32 ( <i>t</i> , $J=8.0$ )	7.24 ( <i>t</i> , $J=7.5$ )
H–C(7)	7.05 ( <i>d</i> , $J=7.8$ )	7.63 ( <i>t</i> , $J=7.8$ )	7.62 ( <i>t</i> , $J=7.8$ )	7.43 ( <i>d</i> , $J=7.8$ )	7.39 ( <i>d</i> , $J=7.5$ )
H–C(8)		7.84 ( <i>d</i> , $J=8.0$ )	7.83 ( <i>d</i> , $J=7.8$ )		
HO–C(8)	10.13 ( <i>br. s</i> )				
H–N(9)	11.75 ( <i>br. s</i> )	12.22 ( <i>br. s</i> )	12.18 ( <i>br. s</i> )	11.48 ( <i>br. s</i> )	10.99 ( <i>br. s</i> )
H–C(14)					5.15–5.19 ( <i>m</i> )
HO–C(14)					5.95 ( <i>d</i> , $J=4.5$ )
Me(15) or CH <sub>2</sub> (15)	2.83 ( <i>s</i> )	2.94 ( <i>s</i> )	2.89 ( <i>s</i> )	2.84 ( <i>s</i> )	3.89–3.94, 3.82–3.87 ( <i>2m</i> )
HO–C(15)					4.86 ( <i>t</i> , $J=5.5$ )
Me(17) or H–N(17)	3.97 ( <i>s</i> )	8.83 ( <i>d</i> , $J=8.0$ )	8.99 ( <i>d</i> , $J=8.0$ )	3.97 ( <i>s</i> )	3.91 ( <i>s</i> )
H–C(18)		4.57–4.61 ( <i>m</i> )	4.32–4.33 ( <i>m</i> )		
CH <sub>2</sub> (19)		2.25–2.31, 2.10–2.18 ( <i>2m</i> )	2.13–2.15, 1.93–1.97 ( <i>2m</i> )		
CH <sub>2</sub> (20)		2.42–2.50 ( <i>m</i> )	2.33–2.39, 2.26–2.29 ( <i>2m</i> )		
HO–C(22)		12.99 ( <i>br. s</i> )			
Me(23)		3.54 ( <i>s</i> )			
H–C(1')				4.94 ( <i>d</i> , $J=7.5$ )	4.96 ( <i>d</i> , $J=7.5$ )
H–C(2')				3.40–3.47 ( <i>m</i> )	3.40–3.47 ( <i>m</i> )
H–C(3')				3.40–3.47 ( <i>m</i> )	3.40–3.47 ( <i>m</i> )
H–C(4')				3.24–3.27 ( <i>m</i> )	3.22–3.27 ( <i>m</i> )
H–C(5')				3.33–3.36 ( <i>m</i> )	3.34–3.37 ( <i>m</i> )
CH <sub>2</sub> (6')				3.77–3.80, 3.51–3.56 ( <i>2m</i> )	3.76–3.80, 3.51–3.55 ( <i>2m</i> )

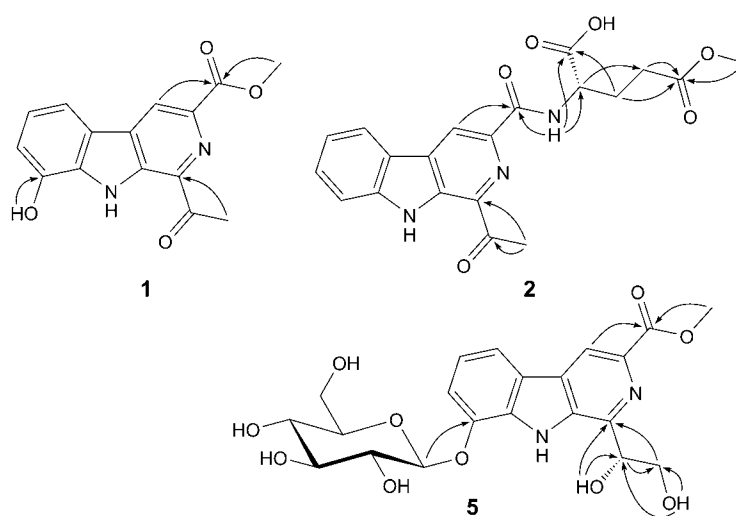
Fig. 2. Key HMBCs (H  $\rightarrow$  C) of compounds **1**, **2**, and **5**

Table 2.  $^{13}\text{C}$ -NMR Spectroscopic Data for the Isolated  $\beta$ -Carboline Derivatives **1–5** (125 MHz,  $(\text{D}_6)$ DMSO,  $\delta$  in ppm). Atom numbering as indicated in Fig. 1.

	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>
C(1)	135.3 ( <i>s</i> )	133.9 ( <i>s</i> )	133.8 ( <i>s</i> )	135.7 ( <i>s</i> )	146.4 ( <i>s</i> )
C(3)	135.7 ( <i>s</i> )	138.0 ( <i>s</i> )	138.6 ( <i>s</i> )	135.9 ( <i>s</i> )	135.7 ( <i>s</i> )
C(4)	121.3 ( <i>d</i> )	118.1 ( <i>d</i> )	117.7 ( <i>d</i> )	121.5 ( <i>d</i> )	116.8 ( <i>d</i> )
C(5)	112.9 ( <i>d</i> )	122.3 ( <i>d</i> )	122.2 ( <i>d</i> )	116.2 ( <i>d</i> )	115.8 ( <i>d</i> )
C(6)	122.3 ( <i>d</i> )	120.9 ( <i>d</i> )	120.7 ( <i>d</i> )	122.1 ( <i>d</i> )	120.9 ( <i>d</i> )
C(7)	114.8 ( <i>d</i> )	129.4 ( <i>d</i> )	129.2 ( <i>d</i> )	115.4 ( <i>d</i> )	114.0 ( <i>d</i> )
C(8)	143.4 ( <i>s</i> )	113.3 ( <i>d</i> )	113.3 ( <i>d</i> )	144.1 ( <i>s</i> )	144.2 ( <i>s</i> )
C(10)	134.8 ( <i>s</i> )	134.9 ( <i>s</i> )	134.8 ( <i>s</i> )	134.9 ( <i>s</i> )	135.0 ( <i>s</i> )
C(11)	131.7 ( <i>s</i> )	131.9 ( <i>s</i> )	131.9 ( <i>s</i> )	132.4 ( <i>s</i> )	131.6 ( <i>s</i> )
C(12)	122.2 ( <i>s</i> )	120.3 ( <i>s</i> )	120.3 ( <i>s</i> )	121.9 ( <i>s</i> )	122.4 ( <i>s</i> )
C(13)	130.8 ( <i>s</i> )	142.4 ( <i>s</i> )	142.3 ( <i>s</i> )	131.6 ( <i>s</i> )	128.5 ( <i>s</i> )
C(14)	201.2 ( <i>s</i> )	200.9 ( <i>s</i> )	200.6 ( <i>s</i> )	200.9 ( <i>s</i> )	74.4 ( <i>d</i> )
C(15)	25.5 ( <i>q</i> )	25.9 ( <i>q</i> )	25.6 ( <i>q</i> )	25.6 ( <i>q</i> )	65.2 ( <i>t</i> )
C(16)	165.2 ( <i>s</i> )	164.2 ( <i>s</i> )	162.9 ( <i>s</i> )	165.1 ( <i>s</i> )	165.9 ( <i>s</i> )
C(17)	52.2 ( <i>q</i> )			52.3 ( <i>q</i> )	52.0 ( <i>q</i> )
C(18)		51.7 ( <i>d</i> )	53.3 ( <i>d</i> )		
C(19)		26.4 ( <i>t</i> )	28.7 ( <i>t</i> )		
C(20)		29.9 ( <i>t</i> )	32.5 ( <i>t</i> )		
C(21)		173.0 ( <i>s</i> )			
C(22)		173.0 ( <i>s</i> )			
C(23)		51.4 ( <i>q</i> )			
C(1')				102.6 ( <i>d</i> )	102.6 ( <i>d</i> )
C(2')				73.2 ( <i>d</i> )	73.4 ( <i>d</i> )
C(3')				77.3 ( <i>d</i> )	77.4 ( <i>d</i> )
C(4')				69.7 ( <i>d</i> )	69.8 ( <i>d</i> )
C(5')				75.7 ( <i>d</i> )	75.9 ( <i>d</i> )
C(6')				60.7 ( <i>t</i> )	60.8 ( <i>t</i> )

was obtained as a yellowish amorphous solid. The HR-ESI-MS, and the  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data provided the molecular formula  $\text{C}_{20}\text{H}_{19}\text{N}_3\text{O}_6$ . In the  $^1\text{H}$ -NMR spectrum, resonances for four mutually coupled, vicinal aromatic H-atoms at ( $\delta(\text{H})$  7.35, 7.63 (*2t*,  $J = 7.8$ ), and 8.46, 7.84 (*2d*,  $J = 8.0$ )) were indicative of the presence of an unsubstituted aromatic ring *A*. A broad *singlet* NH H-atom signal at  $\delta(\text{H})$  12.22 (br. *s*,  $\text{D}_2\text{O}$ -exchangeable) and an aromatic H-atom *singlet* at  $\delta(\text{H})$  9.11 (*s*) were also displayed. These signals were indicative of a  $\beta$ -carboline skeleton. As in compound **1**, a MeO signal at  $\delta(\text{H})$  3.54 (*s*), a Me signal at  $\delta(\text{H})$  2.94 (*s*), together with the C-atom signals at  $\delta(\text{C})$  173.0 and 51.4, and 200.9 and 25.9, also indicated the presence of a COOMe group and an Ac group, respectively. In addition, five mutually coupled H-atom signals at  $\delta(\text{H})$  4.57–4.61 (*m*, 1 H) and 2.50–2.10 (*m*, 4 H), a COOH signal at  $\delta(\text{H})$  12.99 (br. *s*), along with three saturated C-atom signals at  $\delta(\text{C})$  51.7–26.4, and a COOH signal at  $\delta(\text{C})$  173.0, suggested the presence of a  $\text{CHCH}_2\text{CH}_2$  moiety and of a carboxylic acid unit with the aid of HSQC.

The HMBCs between the signals at  $\delta(\text{H})$  2.25–2.31 and 2.10–2.18 ( $\text{CH}_2(19)$ ), and  $\delta(\text{C})$  173.0 (C(22)), between the signals at  $\delta(\text{H})$  3.54 (Me(23)) and  $\delta(\text{C})$  173.0 (C(21)), and between those  $\delta(\text{H})$  8.83 (H–N(17)) and  $\delta(\text{C})$  164.2 (C(16)) and 51.7 (C(18)) led

to the construction of the fragment  $\text{CONHCH}(\text{COOH})\text{CH}_2\text{CH}_2\text{CO}_2\text{Me}$  [7]. Moreover, the  $^3J$  correlations from  $\delta(\text{H})$  2.94 (H–C(15)) to  $\delta(\text{C})$  133.9 (C(1)), and from  $\delta(\text{H})$  9.11 (H–C(4)) to  $\delta(\text{C})$  164.2 (C(16)), evidenced that the side chains Ac and  $\text{CONHCH}(\text{CO}_2\text{H})\text{CH}_2\text{CH}_2\text{CO}_2\text{Me}$  were at C(1) and C(3), respectively (Fig. 2). Compound **2** exhibited a positive optical rotation, similar to dichotomide XII [7]. Thus, the absolute configuration at C(18) was determined as (*S*). On the basis of the above results, the structure of **2** was deduced as shown in Fig. 1, and named dichotomine G.

Compound **3**, a yellowish amorphous solid, had the molecular formula  $\text{C}_{19}\text{H}_{17}\text{N}_3\text{O}_6$  deduced from HR-ESI-MS ( $m/z$  406.1016 ( $[\text{M} + \text{Na}]^+$ )). The  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra displayed very similar signals to those of **2**, except for the absence of a MeO signal in **3**. This is in agreement with a comparison of the molecular formulae of **3** ( $\text{C}_{19}\text{H}_{17}\text{N}_3\text{O}_6$ ) and **2** ( $\text{C}_{20}\text{H}_{19}\text{N}_3\text{O}_6$ ). The absolute configuration at C(18) was also determined to be (*S*), as in compound **2** [7], and the structure of compound **3**, named dichotomine H, was thus established as shown in Fig. 1.

Compound **4**, purified as a pale yellow amorphous solid, was determined to have the molecular formula  $\text{C}_{21}\text{H}_{22}\text{N}_2\text{O}_9$  by HR-ESI-MS ( $m/z$  445.1251 ( $[\text{M} - \text{H}]^-$ )). The  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data of **4** were similar to those of **1**, the major difference being the presence of an additional glucosyl unit ( $\delta(\text{H})$  4.94 (*d*,  $J = 7.5$ , H–C(1')), 3.24–3.80 (*m*, 6 H);  $\delta(\text{C})$  102.6, 77.3, 75.7, 73.2, 69.7, 60.7). Acid hydrolysis of **4** with 1M  $\text{H}_2\text{SO}_4$  furnished a sugar, which was identified as D-glucose by its optical rotation and TLC comparison with an authentic sample. The  $^3J(1,2)$  coupling constant (7.5 Hz) indicated a  $\beta$ -D-glucoside. The sugar unit was connected to C(8) through a C–O linkage as indicated by the HMBC between the signal at  $\delta(\text{H})$  4.94 (H–C(1')) and that at  $\delta(\text{C})$  144.1 (C(8)). Thus, structure **4** was assigned to dichotomine I.

Compound **5** was obtained as a yellowish amorphous solid. On the basis of its HR-ESI-MS ( $m/z$  463.1355,  $[\text{M} - \text{H}]^-$ ), along with the  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data (Tables 1 and 2), its molecular formula was established as  $\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_{10}$ . The  $^1\text{H}$ -NMR spectrum of **5** was found to be very similar to that of **4**, except that the Ac group of **4** was replaced by a 1,2-dihydroxyethyl group ( $\delta(\text{H})$  5.15–5.19 (*m*), 3.89–3.94 (*m*), 3.82–3.87 (*m*),  $\delta(\text{C})$  74.4, 65.2). The HMBCs between the signals at  $\delta(\text{H})$  5.95 (HO–C(14)) and  $\delta(\text{C})$  65.2 (C(15)); the signals at  $\delta(\text{H})$  4.86 (HO–C(15)) and  $\delta(\text{C})$  74.4 (C(14)), and those at  $\delta(\text{H})$  3.89–3.94, 3.82–3.87 (CH<sub>2</sub>(15)) and  $\delta(\text{C})$  146.4 (C(1)) confirmed the above conclusion (Fig. 2).

Acid hydrolysis of **5** with 1M  $\text{H}_2\text{SO}_4$  liberated compound **5a** as the aglycon and D-glucose. The absolute configuration at C(14) in **5** is proposed as (*R*) by comparison of the sign of the specific rotation of **5a** with that of dichotomine C [15]. Thus, structure **5** was assigned to dichotomine J.

Besides compounds **1–5**, the nine known  $\beta$ -carboline alkaloids dichotomides I, III, V, and VII (**6–9**, resp.) [7][15], stellarines A and C (**10–11**, resp.) [16][17], dichotomine B (**12**) [15], glucodichotomine B (**13**) [18], and 1-acetyl-3-carboxy- $\beta$ -carboline (**14**) [19], were also isolated and identified by comparison with spectroscopic data recorded in the literatures.

**Biological Studies.** The cytotoxicities of compounds **1–14** against four human cancer cell lines Bel7402 (human liver cancer), SMMC-7721 (human liver cancer), HCT 116 (human colon cancer), and H460 (human lung cancer) were tested by the MTT (= 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide) method

[20]. Compound **12** showed cytotoxicity only towards SMMC-7721 cells with an  $IC_{50}$  value of 85.36  $\mu\text{M}$ . Compound **13** exhibited cytotoxicity towards HCT116 and SMMC-7721 cells with  $IC_{50}$  values 50.29 and 74.52  $\mu\text{M}$ , respectively. The other isolated compounds showed no or low cytotoxic activities ( $IC_{50}$  values  $> 100 \mu\text{M}$ ) against the tested tumor cells.

This study was supported by the *National Natural Science Foundation of China* (No. 81073009) and the *Priority Academic Program Development of Jiangsu Higher Education Institutions* (PAPD).

### Experimental Part

*General.* All the reagents and solvents were of the anal. grade (*Jiangsu Hanbang Sci. & Tech. Co. Ltd.*, Huaian, P. R. China). Column chromatography (CC): commercial silica gel ( $\text{SiO}_2$ ; 100–200 and 200–300 mesh; *Qingdao Marine Chemical Factory*, Qingdao, P. R. China), *RP-18*  $\text{SiO}_2$  (40–63  $\mu\text{m}$ ; *Fuji Silysia Chemical Ltd.*), *D101* macroporous resin (*The Chemical Plant of Nankai University*, Tianjin, P. R. China), and *Sephadex LH-20* (*Pharmacia, Amersham Biosciences*, Uppsala, Sweden). TLC:  $\text{SiO}_2$  plates; detection by spraying with vanillin/ $\text{H}_2\text{SO}_4$  in EtOH, followed by heating. Optical rotations: *JASCO P-1020* polarimeter. UV Spectra: *Shimadzu UV-2450* spectropolarimeter;  $\lambda_{\text{max}}$  (log  $\epsilon$ ) in nm. IR Spectra: *Bruker Tensor-27* spectrometer; KBr pellets; in  $\text{cm}^{-1}$ . 1D- and 2D-NMR spectra: *Bruker AV-500* spectrometer; at 500 ( $^1\text{H}$ ) and 125 MHz ( $^{13}\text{C}$ ); in ( $\text{D}_6$ )DMSO;  $\delta$  in ppm rel. to TMS as an internal standard,  $J$  in Hz. ESI-MS: *Agilent 1100 Series LC/MSD Trap* mass spectrometer; in  $m/z$ ; HR-ESI-MS: *Micro Q-TOF MS* instrument; in  $m/z$ .

*Plant Material.* The roots of *S. dichotoma* L. var. *lanceolata* BUNGE were purchased from Nanjing Medicine Company, and identified by Prof. Mian Zhang, the Research Department of Pharmacognosy, China Pharmaceutical University. A voucher specimen (No. 20100801) was deposited with the Department of Natural Medicinal Chemistry, China Pharmaceutical University.

*Extraction and Isolation.* The powdered dry roots of *S. dichotoma* var. *lanceolata* (8.0 kg) were extracted four times with 95% aq. EtOH under reflux, each for 2 h. The extract was concentrated *in vacuo*. Then, the residue (500.0 g) was suspended in  $\text{H}_2\text{O}$  and partitioned by precipitation. The supernatant was subjected to CC (*D101*; EtOH/ $\text{H}_2\text{O}$  0:100, 30:70, 50:50, 70:30, 100:0 (v/v)) to yield five fractions, *Frs. 1–5*. *Fr. 2* (30:70, 50.0 g) was subjected to CC (*MCI*; MeOH/ $\text{H}_2\text{O}$  4:6) and further purified by CC (*RP-18* gel; MeOH/ $\text{H}_2\text{O}$  3:7) to affording **12** (50.6 mg). *Fr. 3* (50:50, 20.0 g) was also subjected to CC (*MCI*; MeOH/ $\text{H}_2\text{O}$  30:70  $\rightarrow$  100:0) to furnish four subfractions; *Subfrs. 3.1–3.4*. *Subfr. 3.2* (50:50) was separated by CC ( $\text{SiO}_2$ ;  $\text{CH}_2\text{Cl}_2/\text{MeOH}$  10:1  $\rightarrow$  1:1) to afford seven pooled subfractions, *Subfrs. 3.2.1–3.2.7*. *Subfrs. 3.2.4* (10:3) and *3.2.7* (1:1) were both subjected to CC (*RP-18* gel), followed by CC (*Sephadex LH-20*; MeOH) to afford **5** (11.5 mg), **9** (3.2 mg), and **13** (7.4 mg), resp. *Subfr. 3.3* (70:30) was submitted to CC ( $\text{SiO}_2$ ;  $\text{CH}_2\text{Cl}_2/\text{MeOH}$  4:1  $\rightarrow$  1:1), followed by CC (*RP-18* gel; MeOH/ $\text{H}_2\text{O}$  30:70  $\rightarrow$  70:30), to furnish three subfractions *Subfrs. 3.3.1–3.3.3*. *Subfrs. 3.3.1* (30:70) and *3.3.3* (70:30) were further purified by CC (*Sephadex LH-20*; MeOH) to give **2** (8.1 mg) and **4** (6.7 mg), resp. Prep. TLC of *Subfr. 3.3.2* (50:50) with AcOEt/MeOH 8:2 yielded **3** (14.3 mg;  $R_f$  0.2) and **14** (4.6 mg;  $R_f$  0.5). *Fr. 4* (70:30; 5.0 g) was chromatographed continuously into five subfractions, *Subfrs. 4.1–4.5* using a  $\text{SiO}_2$  CC (gradient of  $\text{CH}_2\text{Cl}_2/\text{MeOH}$ ). *Subfr. 4.3* was successively subjected to CC (*RP-18* gel; MeOH/ $\text{H}_2\text{O}$  40:60  $\rightarrow$  100:0) and CC (*Sephadex LH-20*; MeOH) to give **10** (60.4 mg).

The precipitate (90.0 g) was subjected to CC ( $\text{SiO}_2$ ; PE/AcOEt 100:1, 100:3, 20:1, 10:1, 10:3, 2:1, 1:1, 0:1) to give eight fractions, *Frs. A–H*. *Fr. E* (10:3) was applied to CC (silica gel; petroleum ether (PE)/acetone) to furnish five subfractions, *Subfrs. E.1–E.5*. *Subfr. E.2* was further separated by CC ( $\text{SiO}_2$ ; PE/AcOEt 10:4) to give compound **7** (15.4 mg). *Subfr. E.3* was further purified by CC (*Sephadex LH-20*) to give compounds **1** (7.8 mg) and **8** (4.6 mg). *Fr. F* (2:1) was subjected to CC (*RP-18* gel; MeOH/ $\text{H}_2\text{O}$  50:50  $\rightarrow$  100:0) to give four subfractions, *Subfrs. F.1–F.4*. *Subfr. F.1* was further separated by CC (*Sephadex LH-20*; MeOH) to yield compounds **6** (2.5 mg) and **11** (5.6 mg).

*Dichotomine F* (= *Methyl 1-Acetyl-8-hydroxy-9H- $\beta$ -carboline-3-carboxylate*; **1**). Yellow powder. UV (MeOH): 236 (4.19), 286 (4.20), 385 (3.56). IR: 3373, 2963, 2941, 1702, 1671, 1587, 1437, 1415, 1352, 1259,

1232, 1217. <sup>1</sup>H- and <sup>13</sup>C-NMR: see *Tables 1* and 2, resp. HR-ESI-MS: 307.0686 ( $[M + Na]^+$ , C<sub>15</sub>H<sub>12</sub>N<sub>2</sub>NaO<sub>4</sub><sup>+</sup>; calc. 307.0689).

*Dichotomine G* (= (2S)-2-[(1-Acetyl-9H-β-carbolin-3-yl)carbonylamino]-5-methoxy-5-oxopentanoic Acid = N-[(1-Acetyl-9H-pyrido[3,4-b]indol-3-yl)carbonyl]-L-glutamic Acid 5-Methyl Ester; **2**). Yellowish powder.  $[\alpha]_D^{23} = +28.4$  ( $c = 0.05$ , MeOH). UV (MeOH): 220 (4.25), 286 (4.34), 376 (3.51). IR: 3385, 2952, 1738, 1661, 1537, 1494, 1450, 1335, 1254, 1182. <sup>1</sup>H- and <sup>13</sup>C-NMR: see *Tables 1* and 2, resp. HR-ESI-MS: 396.1199 ( $[M - H]^-$ , C<sub>20</sub>H<sub>18</sub>N<sub>3</sub>O<sub>6</sub><sup>-</sup>; calc. 396.1201).

*Dichotomine H* (= N-[(1-Acetyl-9H-β-carbolin-3-yl)carbonyl]-L-glutamic Acid; **3**). Yellowish powder.  $[\alpha]_D^{23} = +12.0$  ( $c = 0.02$ , MeOH). UV (MeOH): 219 (3.85), 286 (3.87), 377 (3.04). IR: 3389, 1592, 1495, 1410, 1185. <sup>1</sup>H- and <sup>13</sup>C-NMR: see *Tables 1* and 2, resp. HR-ESI-MS: 406.1016 ( $[M + Na]^+$ , C<sub>19</sub>H<sub>17</sub>N<sub>3</sub>NaO<sub>6</sub><sup>+</sup>; calc. 406.1010).

*Dichotomine I* (= Methyl 1-Acetyl-8-(β-D-glucopyranosyloxy)-9H-β-carbolin-3-carboxylate; **4**). Pale yellow powder.  $[\alpha]_D^{23} = -6.6$  ( $c = 0.07$ , MeOH). UV (MeOH): 208 (4.42), 284 (4.08), 376 (3.29). IR: 3772, 3428, 2922, 1717, 1662, 1510, 1433, 1369, 1258, 1101. <sup>1</sup>H- and <sup>13</sup>C-NMR: see *Tables 1* and 2, resp. HR-ESI-MS: 445.1251 ( $[M - H]^-$ , C<sub>21</sub>H<sub>21</sub>N<sub>2</sub>O<sub>5</sub><sup>-</sup>; calc. 445.1253).

*Dichotomine J* (= Methyl 1-[(1R)-1,2-Dihydroxyethyl]-8-(β-D-glucopyranosyloxy)-9H-β-carbolin-3-carboxylate; **5**). Yellowish powder.  $[\alpha]_D^{20} = -17.8$  ( $c = 0.06$ , MeOH). UV (MeOH): 229 (4.14), 268 (4.23), 306 (3.62). IR: 3412, 2963, 2941, 1702, 1671, 1587, 1437, 1415, 1352, 1259, 1232, 1217. <sup>1</sup>H- and <sup>13</sup>C-NMR: see *Tables 1* and 2, resp. HR-ESI-MS: 463.1355 ( $[M - H]^-$ , C<sub>21</sub>H<sub>23</sub>N<sub>2</sub>O<sub>10</sub><sup>-</sup>; calc. 463.1358).

*Acid Hydrolysis of Compounds 4 and 5*. A soln. of **4** or **5** (each 3.0 mg) in 1M H<sub>2</sub>SO<sub>4</sub> (4.0 ml) was heated under reflux for 3 h. After cooling, the mixture was extracted with BuOH three times. The acid aq. layer was neutralized with BaCl<sub>2</sub> to give a BaSO<sub>4</sub> precipitate. After filtering, the aq. layer was concentrated to dryness under reduced pressure. TLC Analysis with authentic glucose as reference (BuOH/AcOH/H<sub>2</sub>O 4:1:5 (v/v/v), upper layer), together with its optical rotation ( $[\alpha]_D^{23} = +52.6$ ,  $c = 0.03$ , H<sub>2</sub>O), indicated the presence of D-glucose. The BuOH layer was washed with brine then dried (MgSO<sub>4</sub>). After removal of the solvent under reduced pressure, the residue was purified CC (*Sephadex LH-20*; MeOH) to yield compounds **1** (1 mg) and **5a** (1.5 mg), resp.

*Methyl 1-[(1R)-1,2-Dihydroxyethyl]-8-hydroxy-9H-β-carbolin-3-carboxylate (5a)*. Yellow powder.  $[\alpha]_D^{25} = -20.9$  ( $c = 0.15$ , MeOH). <sup>1</sup>H-NMR (500 MHz, CD<sub>3</sub>OD): 4.09 (s, MeO-C(16)), 4.01–4.07 (m, CH<sub>2</sub>(15)), 5.51–5.54 (m, H-C(14)), 7.05 (d,  $J = 7.5$ , H-C(6)), 7.21 (t,  $J = 7.5$ , H-C(7)), 7.76 (d,  $J = 7.5$ , H-C(5)), 8.85 (s, H-C(4)).

*Cytotoxicity Assay*. Cells were seeded in 96-well plates 12 h before treatment and continuously exposed to different concentrations of compounds (120, 60, 30, 15, 7.5, 3.25 μM). After 48 h, 20 μl of MTT (= 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide; 5 mg/ml) soln. were added to each well. The old medium was removed after 4 h, and then 100 μl of DMSO was added to each well. The optical density was measured at 570 nm with a *Spectra Shell Microplate Reader* (Tecan, Research Triangle Park, NC, USA). The cells were obtained from the Cell Bank of the Shanghai Institute of Cell Biology. All assays were carried out in triplicate.

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Received December 2, 2011